

Numerical study of the electroporation pulse shape effect on molecular uptake of biological cells

Damijan Miklavcic and Leila Towhidi

Faculty of Electrical Engineering, University of Ljubljana, Ljubljana, Slovenia

Received 16 November 2009

Accepted 2 December 2009

Correspondence to: Prof. Damijan Miklavcic, PhD, Faculty of Electrical Engineering, University of Ljubljana, Trzaska 25, 1000 Ljubljana, Slovenia. Phone: +386 1 4768 456; Fax: +386 1 4264 658; E-mail: damijan.miklavcic@fe.uni-lj.si

Disclosure: No potential conflicts of interest were disclosed.

Background. In order to reduce the side-effects of chemotherapy, combined chemotherapy-electroporation (electrochemotherapy) has been suggested. Electroporation, application of appropriate electric pulses to biological cells, can significantly enhance molecular uptake of cells due to formation of transient pores in the cell membrane. It was experimentally demonstrated that the efficiency of electroporation is under the control of electric pulse parameters. However, the theoretical basis for these experimental results is not fully explained. In order to predict the outcome of experiments and optimize the efficiency of electroporation before each treatment, we developed a model to investigate the effect of pulse shape on efficiency of electroporation.

Results. Our model is based on a developed chemical-kinetics scheme and trapezium barrier model, while self-consistency was taken into account. This model is further supplemented with a molecular transport model to acquire the molecular uptake of cells. The investigated pulse shapes in this study were unipolar rectangular pulses with different rise and fall times, triangular, sinusoidal and bipolar rectangular pulses and also sinusoidal modulated unipolar pulses with different percentages of modulation. The obtained results from our modelling and simulations are in good agreement with previously published experimental results.

Conclusions. We therefore conclude that this model can be used to predict the effects of arbitrarily shaped electroporation pulses on cell membrane conductivity and molecular transport across the cell membrane.

Key words: electrochemotherapy, optimization; membrane permeability; membrane conductivity

Introduction

Cancer is a leading cause of death around the world and for this reason it has drawn the attention of many researchers. Chemotherapy has been used for many years and is one of the most common treatments for cancer. Cytotoxic chemotherapeutic drugs are usually hydrophilic with very low transport through the cell membrane and thus high doses of these drugs are needed for treatment. Therefore, while chemotherapy can be quite effective in treating certain cancers by interfering with the cancer cell's ability to grow or reproduce, chemotherapeutic drugs reach all parts of the body, not just the cancer cells. Because of this, there may be many side-effects such as nausea, blood cell deficiency, fatigue and loss of hair during treatment.¹

Two decades ago, electrochemotherapy was suggested for its use in clinical treatment of localized tumors.² Electrochemotherapy consists of electroporation and chemotherapy. Electroporation is a technique in which permeability of the plasma membrane increases transiently and reversibly with appropriate pulse parameters³⁻⁶ and is nowadays widely used not only in electrochemotherapy,⁷⁻⁹ but also in biotechnology^{10,11} and in medical applications such as gene electrotransfer^{12,13} and transdermal drug delivery.¹⁴ Electroporation in combination with chemotherapy can increase drug delivery into the cells and consequently drug doses and thus side-effects of chemotherapy can be reduced.^{10,11,15,16}

It was experimentally demonstrated in a number of studies that the efficiency of electroporation is under the control of electric pulse parameters

such as pulse amplitude, duration, and shape.¹⁷⁻²² Optimization of electric field parameters for successful electroporation requires time-consuming and costly experiments for different experimental criteria unless an appropriate model for this phenomenon can be suggested. Although the known models were proposed in previous studies²³⁻²⁸, they are still unable to explain the effect of some parameters such as pulse shape, pulse repetition frequency and number of pulses on the molecular uptake enhancement of the cells under exposure.

The suggested mechanism for electroporation consists of structural changes resulting in formation of transient aqueous pores in the cell membrane. In order to reveal the exact mechanisms and dynamics of pore formation and closure and more importantly resealing of the membrane, theoretical models have drawn a great deal of attention.

In our present study, we investigated the effect of pulse shape on the efficiency of electroporation

and, therefore, electrochemotherapy using modelling and simulation. Our model was based on a chemical-kinetics scheme with two types of pores²⁹ which have been recently confirmed.³⁰ We used developed equations with field-dependent rate coefficients in order to obtain the pore distribution on the membrane. Besides, the conductivity of pores was defined based on a trapezium barrier model for the image forces.³¹ A self-consistent set of equations was used to consider all simultaneous changes. This model was supplemented with a molecular transport model for a single cell to acquire the molecular uptake of cells. The investigated pulse shapes in this study were unipolar rectangular pulses with different rise and fall times, triangular, sinusoidal and bipolar rectangular pulses and also sinusoidal modulated unipolar pulses with different percentages of modulation – all previously used in experimental studies.

Material and methods

Model description

Modified chemical-kinetics model for electroporation

When a cell is exposed to an external electric field, the induced transmembrane voltage (ITV) starts to increase based on the Laplace equation which leads to structural changes of the cell membrane. Based on a previously suggested²⁹, and recently confirmed³⁰ kinetic model, in the first step the intact closed lipids (C) transform to tilted lipid headgroups (C1). In the second step, the prepores (P1) are formed and finally, in the last step the final pores (P2) are formed. The sequential reaction can be described by:



The permeability of the P1 state is negligibly small and P2 is predominantly responsible for molecular uptake. Pore formation and closure are denoted by k_i and k_{-i} ($i=1,2,3$) rate coefficients, respectively. For simplicity, the rate coefficients k_1 , k_2 and k_3 are considered equal ($k_1=k_2=k_3=k_p$).²⁹ The governed rate laws of constituting steps for the scheme [1] are:

$$\begin{aligned} \frac{d[C(\vec{r}, t)]}{dt} &= -k_p [C(\vec{r}, t)] + k_{-1} [C1(\vec{r}, t)] \\ \frac{d[C1(\vec{r}, t)]}{dt} &= -k_p ([C1(\vec{r}, t)] - [C(\vec{r}, t)]) - k_{-1} [C1(\vec{r}, t)] + k_{-2} [P1(\vec{r}, t)] \\ \frac{d[P1(\vec{r}, t)]}{dt} &= -k_p ([P1(\vec{r}, t)] - [C1(\vec{r}, t)]) - k_{-2} [P1(\vec{r}, t)] + k_{-3} [P2(\vec{r}, t)] \\ \frac{d[P2(\vec{r}, t)]}{dt} &= k_p [P1(\vec{r}, t)] - k_{-3} [P2(\vec{r}, t)] \end{aligned} \quad [2]$$

where t and \vec{r} denote time and position, respectively. [C], [C1], [P1] and [P2] show normalized distribution of each membrane lipid state relative to the initial value of the closed state $[C(\vec{r}, 0)]$.²⁹

Regarding the Van't Hoff relationship in electro-thermodynamics, the rate coefficient of pore formation can be obtained from:^{29,31}

$$k_p = k_{p0} \exp\left(\frac{\Delta V_p \varepsilon_0 (\varepsilon_W - \varepsilon_L)}{2k_B T d_m^2} ITV^2\right) \quad [3]$$

where ITV is the potential difference between the outer and inner layer of the membrane, ΔV_p is the mean volume change due to pore formation, ε_0 is the permittivity of the vacuum and ε_W and ε_L are dielectric constants of water and lipids, respectively. k_B is the Boltzmann constant, d_m is the thickness of the membrane and T is temperature. While the pore formation rate coefficient k_p is electric field-dependent, the closure rate coefficients (k_{-1} , k_{-2} and k_{-3}) are constant and independent of electric field strength.²⁹

Whenever electroporation occurs, an increase in conductivity during the pulse is observed³² which can be explained by the formation of pores in the cell membrane. Based on the trapezium barrier model for the image forces, the intrinsic pore conductivities $\sigma_{p,i}$ ($i=1$ and 2 represents P1 and P2 pores, respectively) are expressed as follows:^{31,33}

$$\sigma_{p,i} = \sigma_{p,i}^0 \exp\left(\alpha_{p,i} n \left| ITV \right| \frac{F}{RT}\right) \quad [4]$$

where

$$\sigma_{p,i}^0 = \frac{\sigma_{ex} + \sigma_{in}}{2} \exp\left(\frac{-\varphi_{im,i}^0 F}{RT}\right) \quad \text{and} \quad \alpha_{p,i} = 1 - \frac{RT}{F\varphi_{im,i}^0} \quad [5]$$

In the above equations, σ_{ex} and σ_{in} are the extracellular and intracellular conductivities respectively, n is the geometrical parameter of the trapezium model for energy barrier, F is the Faraday constant and $\varphi_{im,i}^0$ is the intrinsic pore barrier potential.

Therefore, conductivity of the membrane (σ_m) can be obtained by:

$$\sigma_m(\vec{r}, t) = \sigma_{m0} + [P1(\vec{r}, t)] \times \sigma_{p,1} + [P2(\vec{r}, t)] \times \sigma_{p,2} \quad [6]$$

where σ_{m0} is the physiological/baseline conductivity of the membrane. Thus conductivity at each point on the membrane changes with time during and after the pulse, depending on pore distribution variations which affect ITV and in turn the distribution of pores.

Transmembrane molecular transport model

Based on previous studies³⁴⁻³⁶, we defined two distinct phases for the electroporated membrane and two related transport mechanisms: the first one is the porated phase [P2] with relatively fast relaxation due to pore closure according to Eq. [1]. The second phase is the memory phase [M] due to enhanced membrane perturbation and ruffling with quite slow relaxation³⁴⁻³⁶ which returns to its baseline value with a dual exponential decay function:²⁹

$$[M] = [P2]_e (B \exp(-k_f t) + (1 - B) \exp(-k_s t)) \quad [7]$$

where $[P2]_e$ is the normalized distribution of [P2] pores at the end of the pulse, k_f and k_s are decay rate coefficients for this second phase and B is a constant.

The considered transport mechanisms for these two phases were interactive diffusion through the pores and endocytotic-like transport through the permeabilized area of the membrane.³⁴⁻³⁷ Thus, the permeability of the membrane can be written as the sum of two distinct contributions:

$$P_m(\vec{r}, t) = ([P2(\vec{r}, t)] D_p / d_m) + ([M(\vec{r}, t)] D_r / d_m) \quad [8]$$

where D_p and D_r are the attributed diffusion coefficients for interactive transport and endocytotic-like transport, respectively.

While the membrane is being permeabilized due to the electric field, the molecules pass through the membrane due to a concentration gradient. A quantitative description of diffusion is contained in Fick's first law. The total flux can be approximated by $j = P_m (c^{out} - c^{in})$, where c^{out} and c^{in} are the outer and inner concentrations adjacent to the membrane. The total number of molecules transported through the mem-

TABLE 1. Values of parameters used in simulations

| Parameter | Symbol | Value | Ref. |
|--|----------------|---------------------------------|--------|
| Membrane thickness | d_m | 5e-9 m | 29 |
| Extracellular conductivity | σ_{ex} | 0.14 S/m ^o | 29, 38 |
| Intracellular conductivity | σ_{in} | 0.3 S/m ^b | 29, 39 |
| Initial conductivity of membrane | σ_{m0} | 5e-7 S/m | 40 |
| Extracellular permittivity | ϵ_o | 7.1e-10 As/Vm | 29 |
| Intracellular permittivity | ϵ_i | 7.1e-10 As/Vm | 29 |
| Membrane permittivity | ϵ_m | 4.4e-11 As/Vm ^c | 29 |
| Water relative dielectric constant | ϵ_w | 80 As/Vm | 29 |
| Lipid relative dielectric constant | ϵ_l | 2 As/Vm | 29 |
| Free diffusion coefficient | D_0 | 5e-10 m ² /s | 29 |
| Zero-field equilibrium constant | K_0 | 2e-2 | 29 |
| Mean average aqueous pore volume | ΔV_p | 9e-27 m ³ | 29 |
| Intrinsic barrier potential of P1 state | ϕ_{im1}^0 | 0.13 V | 31 |
| Intrinsic barrier potential of P2 state | ϕ_{im2}^0 | 0.084 V | 31 |
| A geometrical parameter | n | 0.12 | 33 |
| Decay rate coefficient for C1 | k_1 | 10 ⁵ s ⁻¹ | 41-43 |
| Decay rate coefficient for P1 pores | k_2 | 2000 s ⁻¹ | 41-43 |
| Decay rate coefficient for P2 pores | k_3 | 2 s ⁻¹ | 41 |
| Decay rate coefficients for endocytotic-like process | k_r, k_s | 0.044, 0.003 s ⁻¹ | 29 |

^o This is for SMEM. The range of extracellular medium is quite large. ^b Reported between (0.2-0.55) S/m ^c Reported between (4.4-5)*10⁻¹¹ As/Vm

brane (N) was computed with integration of transported molecules through the cell membrane over time and the cell surface:

$$N = N_A \int_{t=0}^{\tau} \int_S j dS dt \quad [9]$$

where S is the surface of the cell membrane, τ is the time at which the quantity of transported molecules is to be determined and N_A is Avogadro's number.

Construction of the model

The simulations in this study were performed using the COMSOL 3.3 package (COMSOL Inc., Burlington, MA) based on the finite element method. To construct the geometrical model, a spherical cell with radius of 5.6 μm was located between two virtual electrodes. Since incorporating an extremely thin membrane is problematic in meshing and solving the problem, we assigned the boundary condition to the membrane.³⁸ We neglected the resting transmembrane voltage. The initial intracellular and extracellular concentrations of probe were set to 0 and 10 mM, respectively. The diffusion coefficients for interactive diffusion and for an

induced endocytotic-like process are considered as $D_0/5$ and $D_0/10000$. These two values, however, depend considerably on the type and size of the transported molecules. The necessary parameters used in our simulations are given in Table 1. Our simulation was designed to solve the Laplace equation considering all related equations in this model (Eq. [2], [3], [6]) taking into account self-consistency of parameters to find the distribution of pores on the cell membrane, spatially and temporally, and all related parameters such as ITV, cell membrane conductivity and permeability. Afterwards, the uptake of the cells for each different pulse shape was obtained. All simulations were performed on a PC (2.8 GHz Pentium IV processor, 3 GB RAM) and

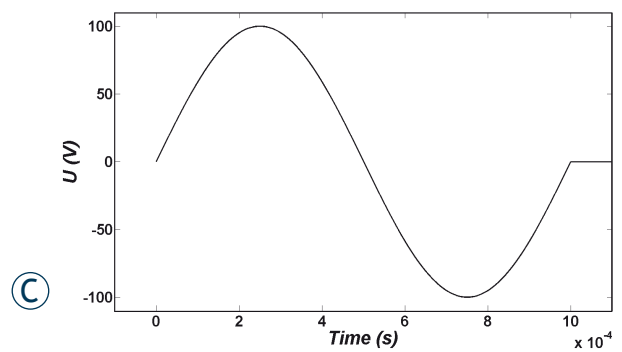
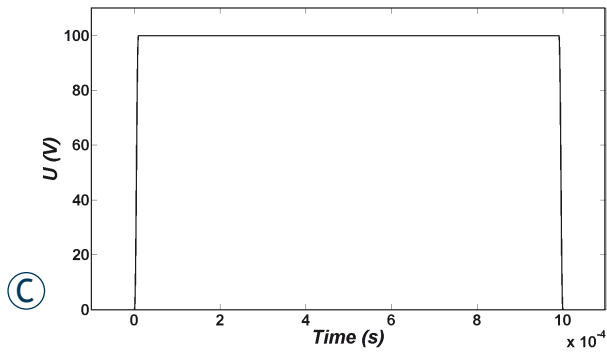
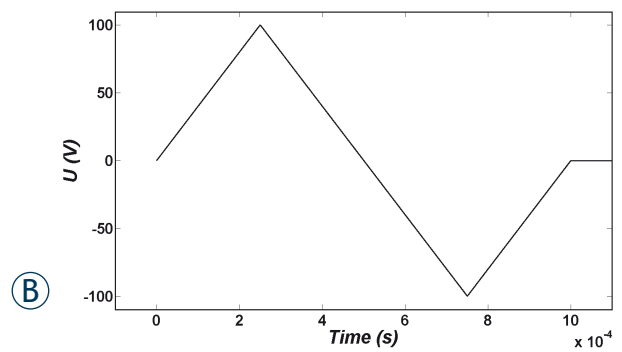
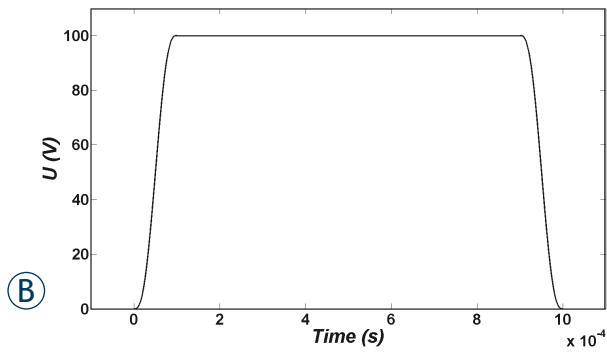
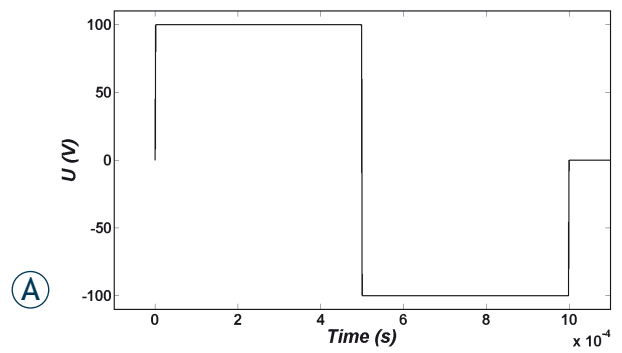
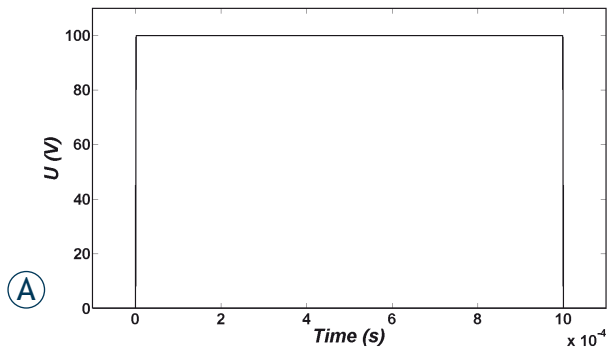


FIGURE 1. The investigated rectangular pulse shapes in this study with rise and fall times of (A) 2, (B) 10 and (C) 100 μ s, respectively.

FIGURE 2. The (A) bipolar rectangular, (B) triangular and (C) sinusoidal pulses considered in this study.

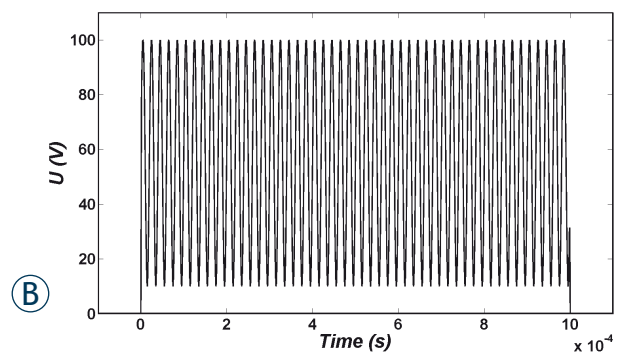
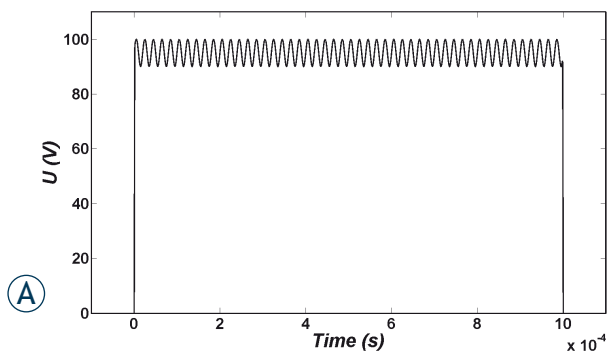


FIGURE 3. The sine-modulated 50 kHz unipolar pulses with (A) 10% and (B) 90% modulation investigated in this paper.

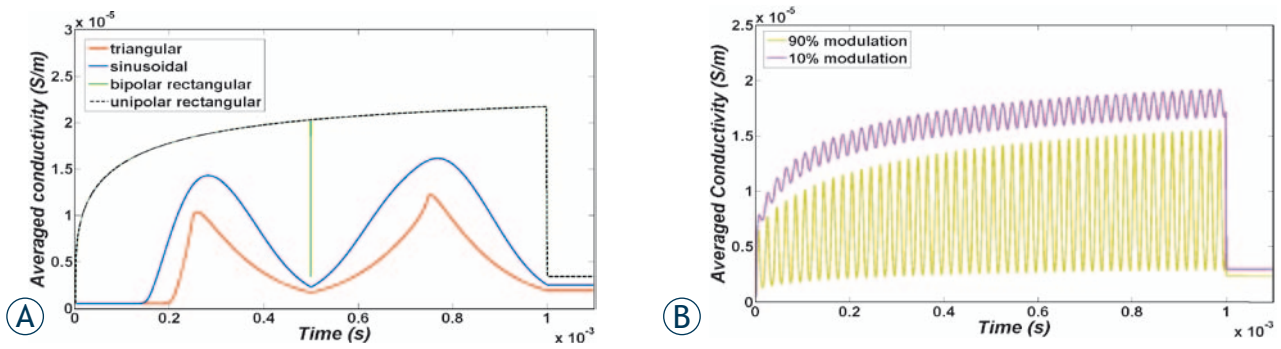


FIGURE 4. Temporal evolution of the overall membrane conductivity during the pulse for (A) unipolar and bipolar rectangular, triangular and sinusoidal pulses and (B) 10% and 90% sine-modulated unipolar pulses.

each simulation lasted 3–25 minutes depending on the considered pulse shape and number of pulses in each train of pulse.

The investigated pulse shapes in this study were unipolar rectangular pulses with different rise and fall times of 2, 10 and 100 μs (Figure 1); triangular, sinusoidal and bipolar rectangular pulses (Figure 2); and also sinusoidal modulated unipolar pulses with different percentages of modulation of 10% and 90% with 50 kHz frequency (Figure 3).

Results

Immediately after the smoothed step pulse is switched on, ITV starts to increase based on the Laplace equation and causes membrane structural changes initiation, which in turn results in the membrane conductivity increase according to Eq. [6]. The temporal behaviour of average conductivities over the cell membrane due to application of the considered pulse shapes (Figures 1, 2 and 3) are shown in Figure 4. All pulses were considered to have a peak of 1 kV/cm and total duration of 1 ms.

It can be observed in Figure 4 that the overall conductivity changes for unipolar and bipolar pulses have negligible differences. The reason for this fact is a very quick switch between positive and negative voltage, as well as ignoring the resting voltage in this model. Besides, a comparison between conductivity increases due to rectangular, triangular and sinusoidal pulses was performed. Figure 4A shows that the largest and smallest changes were due to rectangular and triangular pulse shapes, respectively. Figure 4B shows that a conductivity change due to 10% modulation is higher than the 90% one but both are still lower than the rectangular pulse.

The temporal behaviour of averaged cell membrane permeability for pulses in Figures 1 to 3 is illustrated in Figure 5. Permeability changes occur slowly. Therefore, for bipolar pulses and modulated pulses in which the fall and rise is very fast, there is not enough time for resealing of permeability which causes different behaviour for membrane permeability related to membrane conductivity. Based on Figure 5, we expect the order of efficiency of pulses of the same peak as follows: unipolar

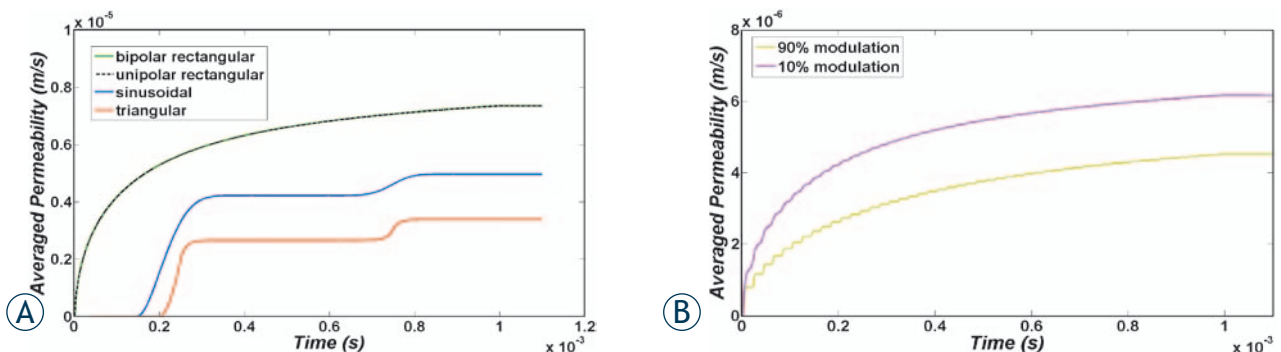


FIGURE 5. Temporal evolution of the overall membrane permeability during the pulse for (A) unipolar and bipolar rectangular, triangular and sinusoidal pulses and (B) 10% and 90% sine-modulated unipolar pulses.

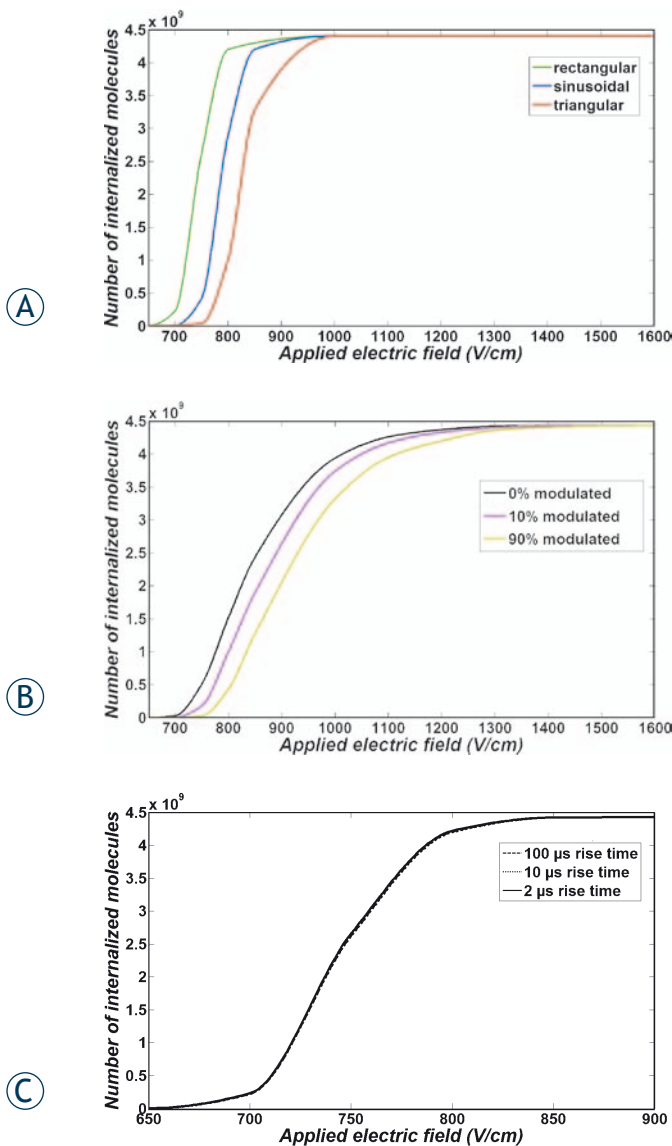


FIGURE 6. Dye uptake 16 minutes after pulse cessation versus electric field amplitude for (A) 8 pulses of 1 ms and 1 Hz unipolar and bipolar rectangular, triangular and sinusoidal pulses, (B) one 1 ms pulse of 0%, 10% and 90% sine-modulated and (C) 8 pulses of 1 ms rectangular pulses with rise and fall times of 2, 10 and 100 μ s.

and bipolar rectangular, 10% modulated, sine, 90% modulated and finally bipolar triangular pulses. Note that unlike membrane conductivity in Figure 4, membrane permeability does not recover as fast after the pulse ceases.

To be able to check the validity of our simulation results, the uptake enhancement of the cell was calculated for the same pulse parameters of previously obtained experimental results.¹⁹ The chosen parameters were 8 pulses of 100 μ s duration and 1 Hz pulse repetition frequency with different pulse strengths for each pulse shape.

Figure 6A shows the results of simulation for 8 pulses of bipolar rectangular, sine and triangular pulses. It shows that the rectangular pulses are more efficient than sinusoidal pulses which in turn are more efficient than triangular pulses. These results are in good agreement with experimentally obtained results.¹⁹

As can be seen in Figure 2A for the bipolar pulses, the pulse switch from positive to negative takes place very fast. During the switch time, the pore creation rate and, therefore, membrane conductivity decrease. But due to very short time of switching related to pulse duration, these changes are negligible in comparison to the conductivity change related to the unipolar pulse (Figure 4A). Consequently, the uptake due to unipolar pulses is larger than bipolar pulses but this difference is negligible and not observable (data not shown). While our simulation shows no significant difference between these two pulse types, in experimental results bipolar pulses are significantly more efficient than unipolar pulses. The reason for this inconsistency is most probably due to neglecting resting voltage in the simulations.

In addition, Figure 6B demonstrates the comparison between unipolar pulses of 0, 10 and 90% modulation. The results are also in good qualitative agreement with previously obtained experimental results.¹⁹ The uptake enhancement results for 8 pulses of unipolar trapezoidal pulses of 1 ms duration with 2, 10 and 100 μ s rise and fall times are shown in Figure 6C. It can be seen from the figure that there is no significant difference between these pulses which is again in good agreement with previously published experimental results.¹⁹

Conclusions

The described model enables determination and prediction of all electrical and diffusion parameters for different pulse shapes. Thus, knowing electrical and diffusion properties of the cells and the specific dye, optimization of the electroporation protocol can be performed before the treatment. Our results show that rectangular pulses are more effective than the sinusoidal and triangular pulses. Besides, our results indicate that the higher the percentage of unipolar pulses modulation with sine shape pulses of 50 kHz, the lower the uptake enhancement of the cells. Moreover, the rise and fall times of unipolar rectangular pulses do not significantly affect the uptake of molecules by the cells. Our simulation results are consistent with experimental observations.

Acknowledgements

This research was supported by the Research Agency of the Republic of Slovenia.

References

- Airley R. *Cancer Chemotherapy: Basic Science to the Clinic*, West Sussex: Wiley-Blackwell; 2009.
- Mir LM, Belehradek M, Domenge C, Orlowski S, Poddevin B, Belehradek J Jr, et al. Electrochemotherapy, a new antitumor treatment: first clinical trial. *CR Acad Sci III* 1991; **313**: 613-8.
- Tsong TY. Electroporation of cell membranes. *Biophys J* 1991; **60**: 297-306.
- Neumann E, Kakorin S, Toensing K. Fundamentals of electroporative delivery of drugs and genes. *Bioelectrochem Bioenergy* 1999; **48**: 3-16.
- Teissie J, Eynard N, Gabriel B, Rols MP. Electroporation of cell membranes. *Adv Drug Del Rev* 1999; **35**: 3-19.
- Neumann E, Kakorin S, Tönsing K. Fundamentals of electroporative delivery of drugs and genes. *Bioelectrochem Bioenergy* 1999; **48**: 3-16.
- Mir LM. Bases and rationale of the electrochemotherapy. *EJC Suppl* 2006; **4**: 38-44.
- Sersa G, Miklavcic D, Cemazar M, Rudolf Z, Pucihar G, Snoj M. Electrochemotherapy in treatment of tumours. *EJSO* 2008; **34**: 232-40.
- Zupanic A, Corovic S, Miklavcic D. Optimization of electrode position and electric pulse amplitude in electrochemotherapy. *Radiol Oncol* 2008; **42**: 93-101.
- Faurie C, Golzio M, Phez E, Teissie J, Rols MP. Electric field induced cell membrane permeabilization and gene transfer: theory and experiments. *Eng Life Sci* 2005; **5**: 179-86.
- Teissie J, Eynard N, Vernhes MC, Bénichou A, Ganeva V, Galutzov B, et al. Recent biotechnological developments of electropulsation. A prospective review. *Bioelectrochem* 2002; **55**: 107-12.
- Golzio M, Mazzolini L, Moller P, Rols MP, Teissie J. Inhibition of gene expression in mice muscle by in vivo electrically mediated siRNA delivery. *Gene Ther* 2005; **12**: 246-51.
- Mesojednik S, Kamensek U, Cemazar M. Evaluation of shRNA-mediated gene silencing by electroporation in LPB fibrosarcoma cells. *Radiol Oncol* 2008; **42**: 82-92.
- Pliquett U, Weaver JC. Feasibility of an electrode-reservoir device for transdermal drug delivery by noninvasive skin electroporation. *IEEE Trans Biomed Eng* 2007; **54**: 536-8.
- Gothelf A, Mir LM, Gehl J. Electrochemotherapy: results of cancer treatment using enhanced delivery of bleomycin by electroporation. *Cancer Treat Rev* 2003; **29**: 371-8.
- Pavselj N, Miklavcic D. Numerical modeling in electroporation-based biomedical applications. *Radiol Oncol* 2008; **42**: 159-68.
- Teissie J, Escoffre JM, Rols MP, Golzio M. Time dependence of electric field effects on cell membranes. A review for a critical selection of pulse duration for therapeutical applications. *Radiol Oncol* 2008; **42**: 196-206.
- Rols MP, Teissie J. Electroporation of mammalian cells to macromolecules: control by pulse duration. *Biophys J* 1998; **75**: 1415-23.
- Kotnik T, Pucihar G, Rebersek M, Mir LM, Miklavcic D. Role of pulse shape in cell membrane electroporation. *Biochim Biophys Acta* 2003; **1614**: 193-200.
- Pucihar G, Mir LM, Miklavcic D. The effect of pulse repetition frequency on the uptake into electroporated cells in vitro with possible applications in electrochemotherapy. *Bioelectrochem* 2002; **57**: 167-72.
- Macek-Lebar A, Sersa G, Kranjc S, Groselj A, Miklavcic D. Optimisation of pulse parameters in vitro for in vivo electrochemotherapy. *Anticancer Res* 2002; **22**: 1731-6.
- Macek-Lebar A, Miklavcic D. Cell electroporation: control by pulse parameters. *Radiol Oncol* 2001; **35**: 193-202.
- DeBruin KA, Krassowska W. Modeling Electroporation in a Single Cell. II. Effects of Ionic Concentrations. *Biophys J* 1999; **77**: 1225-33.
- Gowrishankar TR, Weaver JC. An approach to electrical modeling of single and multiple cells. *Proc Natl Acad Sci USA* 2003; **100**: 3203-8.
- Joshi RP, Hu Q, Schoenbach KH, Bebe SJ. Simulations of electroporation dynamics and shape deformations in biological cells subjected to high voltage pulses. *IEEE Trans Plasma Sci* 2002; **30**: 1536-46.
- Joshi RP, Hu Q, Schoenbach KH. Modeling studies of cell response to ultrashort, high-intensity electric fields—implications for intracellular manipulation. *IEEE Trans Plasma Sci* 2004; **32**: 1677-86.
- Neu JC, Krassowska W. Asymptotic model of electroporation. *Phys Rev E* 1999; **59**: 3471-82.
- Kotnik T, Bobanovic F, Miklavcic D. Sensitivity of transmembrane voltage induced by applied electric fields – a theoretical analysis. *Bioelectrochem Bioenergy* 1997; **43**: 285-91.
- Neumann E, Toensing K, Kakorin S, Budde P, Frey J. Mechanism of electroporative dye uptake by mouse B cells. *Biophys J* 1998; **74**: 98-108.
- Böckmann RA, Groot BL, Kakorin S, Neumann E, Grubmüller H. Kinetics, statistics, and energetics of lipid membrane electroporation studied by molecular dynamics simulations. *Biophys J* 2008; **95**: 1837-50.
- Kakorin S, Neumann E. Ionic conductivity of electroporated lipid bilayer membranes. *Bioelectrochem* 2002; **56**: 163-6.
- Pavlin M, Leben V, Miklavcic D. Electroporation in dense cell suspension – Theoretical and experimental analysis of ion diffusion and cell permeabilization. *Biochim Biophys Acta* 2007; **1770**: 12-23.
- Schmeer M, Seipp T, Pliquett U, Kakorin S, Neumann E. Mechanism for the conductivity changes caused by membrane electroporation of CHO cell – pellets. *Phys Chem Chem Phys* 2004; **6**: 5564-74.
- Glogauer M, Lee W, McCulloch CA. Induced endocytosis in human fibroblasts by electrical fields. *Exp Cell Res* 1993; **208**: 232-40.
- Rols MP, Femenia P, Teissie J. Long-lived macropinosomes take place in electroporated mammalian cells. *Biochem Biophys Res Commun* 1995; **208**: 26-38.
- Zimmermann U, Schnettler R, Klöck G, Watzka H, Donath E, Glaser RW. Mechanisms of electrostimulated uptake of macromolecules into living cells. *Naturwissenschaften* 1990; **77**: 543-5.
- Puc M, Kotnik T, Mir LM, Miklavcic D. Quantitative model of small molecules uptake after in vitro cell electroporation. *Bioelectrochem* 2003; **60**: 1-10.
- Pucihar G, Kotnik T, Valic B, Miklavcic D. Numerical determination of transmembrane voltage induced on irregularly shaped cells. *Annals Biomed Eng* 2006; **34**: 642-52.
- Kotnik T, Miklavcic D, Slivnik T. Time course of transmembrane voltage induced by time-varying electric fields – a method for theoretical analysis and its application. *Bioelectrochem Bioenergy* 1998; **45**: 3-16.
- Pucihar G, Kotnik T, Miklavcic D, Teissie J. Kinetics of transmembrane transport of small molecules into electroporated cells. *Biophys J* 2008; **95**: 2837-48.
- Gowrishankar TR, Pliquett U, Lee RC. Dynamics of membrane sealing in transient electroporation of skeletal muscle membranes. *Ann NY Acad Sci* 1999; **888**: 195-210.
- Hibino M, Itoh H, Kinoshita K. Time courses of cell electroporation as revealed by submicrosecond imaging of transmembrane potential. *Biophys J* 1993; **64**: 1789-800.
- Chernomordik LV, Sukharev SI, Popov SV, Pastushenko VF, Sokirko AV, Abidor IG, et al. The electrical breakdown of cell and lipid membranes: the similarity of phenomenologies. *Biochim Biophys Acta* 1987; **902**: 360-73.